

Loss-of-function mutations in *LEMD3* result in osteopoikilosis, Buschke-Ollendorff syndrome and melorheostosis

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Osteopoikilosis, Buschke-Ollendorff syndrome (BOS) and melorheostosis are disorders characterized by increased bone density¹. The occurrence of one or more of these phenotypes in the same individual or family suggests that these entities might be allelic²⁻⁴. We collected data from three families in which affected individuals had osteopoikilosis with or without manifestations of BOS or melorheostosis. A genome-wide linkage analysis in these families, followed by the identification of a microdeletion in an unrelated individual with these diseases, allowed us to map the gene that is mutated in osteopoikilosis. All the affected individuals that we investigated were heterozygous with respect to a loss-of-function mutation in *LEMD3* (also called *MANT*), which encodes an inner nuclear membrane protein. A somatic mutation in the second allele of *LEMD3* could not be identified in fibroblasts from affected skin of an individual with BOS and an individual with melorheostosis. *XMAN1*, the *Xenopus laevis* ortholog, antagonizes BMP signaling during embryogenesis⁵. In this study, *LEMD3* interacted with BMP and activin-TGF β receptor-activated Smads and antagonized both signaling pathways in human cells.

Osteopoikilosis (OMIM 166700) is an autosomal dominant skeletal dysplasia characterized by a symmetric but unequal distribution of multiple hyperostotic areas in different parts of the skeleton (Fig. 1)⁶. These lesions, usually detected incidentally, represent foci of old remodeled bone with lamellar structure, either connected to adjacent trabeculae of spongy bone or attached to the subchondral cortex⁷. Osteopoikilosis can occur either as an isolated anomaly or in association with other abnormalities of skin and bone. BOS (OMIM 166700),

an autosomal dominant disorder, refers to the association of osteopoikilosis with disseminated connective-tissue nevi. Both elastic-type nevi (juvenile elastoma) and collagen-type nevi (dermatofibrosis lenticularis disseminata) have been described in BOS⁸. Skin or bony lesions can be absent in some family members, whereas other relatives may have both⁹. The co-occurrence of osteopoikilosis and melorheostosis in the same family has been reported in a few instances²⁻⁴. Melorheostosis (OMIM 155950) is characterized by a 'flowing' (rheos) hyperostosis of the cortex of tubular bones. These lesions are usually asymmetric: they may involve only one limb or correspond to a particular sclerotome. They are often accompanied by abnormalities of adjacent soft tissues, such as joint contractures, sclerodermatous skin lesions, muscle atrophy, hemangiomas and lymphoedema^{10,11}.

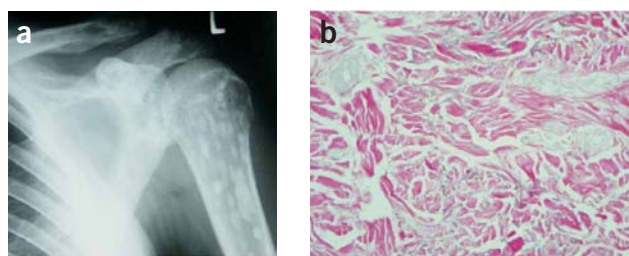


Figure 1 Osteopoikilosis lesions and elastic-type nevus in individual III-3 of family A. (a) Anteroposterior radiograph of the left shoulder showing multiple osteopoikilosis lesions, best visible in the left humerus. (b) Light micrograph of the elastic-type nevus stained with Van Gieson. Original magnification, $\times 100$. Thick and coarse collagen bundles with numerous broad and irregular elastic fibers are present in the mid-dermis.

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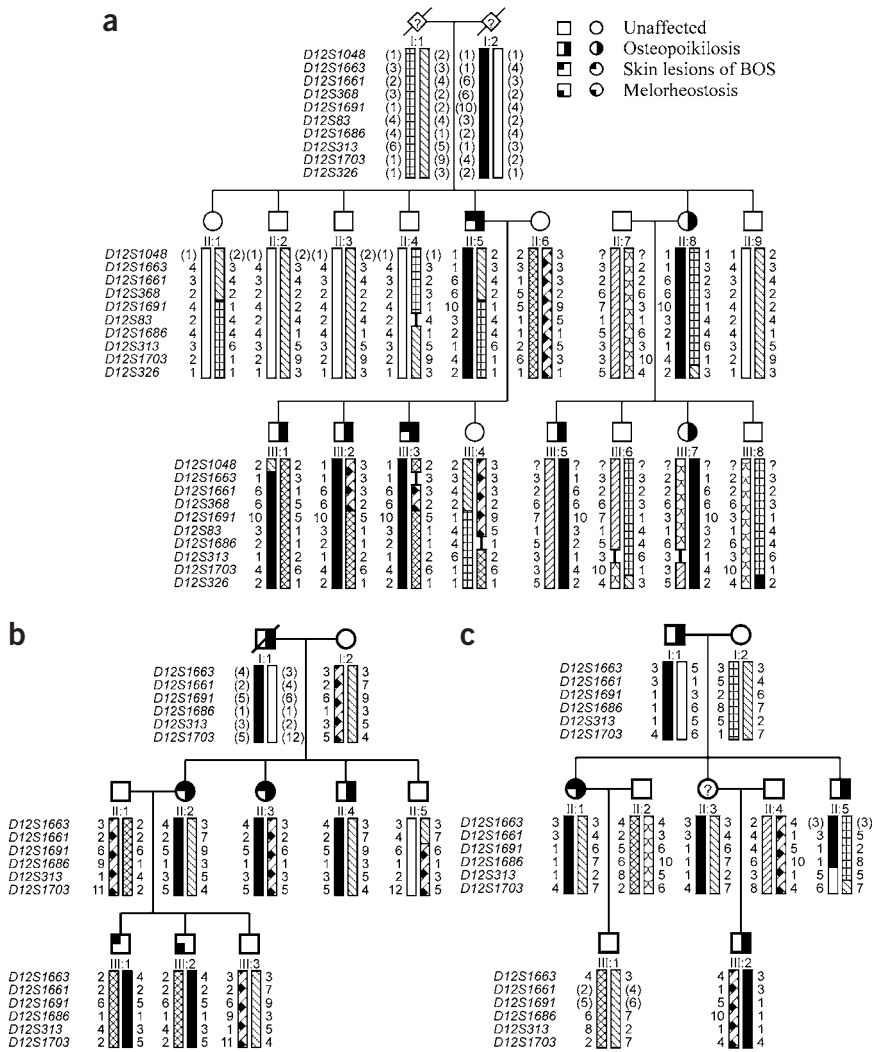


Figure 2 Pedigree structure and haplotypes of the three families with osteopoikilosis, family A (a), family B (b) and family C (c). Inferred alleles are shown in brackets. The haplotype cosegregating with the disorder is indicated with a black bar.

Affected individuals may be asymptomatic or may complain of chronic pain in the affected limb¹².

To unravel the genetic defect of osteopoikilosis, we started a genome-wide linkage analysis in family A (Fig. 2a). Screening of 400 markers with an average spacing of 10 cM resulted in a significant positive lod score (3.744) for two consecutive markers on chromosome 12q13: *D12S368* and *D12S83*. The results of the linkage analysis in the two other families, families B and C (Fig. 2b,c), were consistent with the linkage data obtained in family A. The centromeric boundary of the genetic interval was determined by a recombination event between *D12S1048* and *D12S1663* in individual III-1 (of family A). The telomeric boundary was defined by a recombination event between *D12S1686* and *D12S313* in individual II-5 (of family C), resulting in a candidate region of 23.55 cM on chromosome 12q12–12q14.3. We calculated a combined maximum two-point lod score of 6.691 at $\theta=0$ for markers *D12S1661* and *D12S1691* (Supplementary Table 1 online).

The next step in the genetic analysis was the identification of individual G03-1858, who is affected with proportionate short stature, microcephaly, learning disabilities, ectopic kidneys and

osteopoikilosis. We hypothesized that this individual might have a microdeletion, resulting in the loss of several contiguous genes, including the gene mutated in osteopoikilosis. We investigated this individual for loss of heterozygosity in the candidate region on 12q12–12q14.3 and found loss of heterozygosity for marker *D12S1686*, located in the telomeric part of the interval. The testing of additional markers confirmed the presence of a microdeletion with a centromeric boundary at marker *D12S329* (Fig. 3). The telomeric boundary of the microdeletion was defined at the single-nucleotide polymorphism tsc0527430 using the GeneChip Mapping 10K Array (results for whole genome in Supplementary Fig. 1 online; results for chromosome 12 in Fig. 4c). We then tested more markers in the region of overlap between the microdeletion and the linkage interval in family C, which allowed us to narrow the linkage interval and define a 3.07-Mb critical region for association with osteopoikilosis between marker *D12S329* and μ SAT12.10. This region contains 23 known genes (National Center for Biotechnology Information genome viewer; Fig. 3).

Two of these genes, *WIF1* (Wnt inhibitory factor 1) and *LEMD3* (LEM domain-containing 3), are good candidates for involvement in osteopoikilosis. *WIF1* is involved in Wnt signaling, and *LEMD3* functions in BMP signaling, two pathways important in bone development^{13,14}. Mutation analysis of *WIF1* did not identify any abnormalities in the affected individuals. Sequencing of *LEMD3* identified loss-of-function mutations in all affected individuals of the three families and in three unrelated individuals with osteopoikilosis (Table 1 and Fig. 4e). The splice-site mutation in individual G03-2881 caused skipping of exon 6, resulting in a frameshift and premature stop codon in exon 7 at position 2,021 (Fig. 4d). The deletion of one of the *LEMD3* alleles in individual G03-1858 was corroborated by fluorescence *in situ* hybridization (FISH) analysis with locus-specific probes (Fig. 4b). Re-evaluation of the karyotype (550-band level) was suggestive of, but not conclusive for, the presence of a deletion in the 12q14–15 region (Fig. 4a).

Some reports have suggested that the asymmetric distribution of skin lesions in BOS and the segmental involvement usually observed in melorheostosis result from a somatic mutation^{4,15}. To investigate this possibility, we took skin biopsy samples from two affected individuals, one from an elastic-type nevus in individual III-3 (of family A) with BOS and a second from a hard sclerodermic-like lesion in individual III-2 (of family B) with melorheostosis. Sequence analysis of *LEMD3* on genomic DNA extracted from both skin lesions showed no evidence for an additional somatic mutation in *LEMD3* (or 'second hit'). Analysis of intragenic polymorphisms showed no loss of heterozygosity or allelic imbalance and therefore excluded the possible existence of a partial gene deletion as a somatic mutation. In cDNA from normal and affected skin of the individual

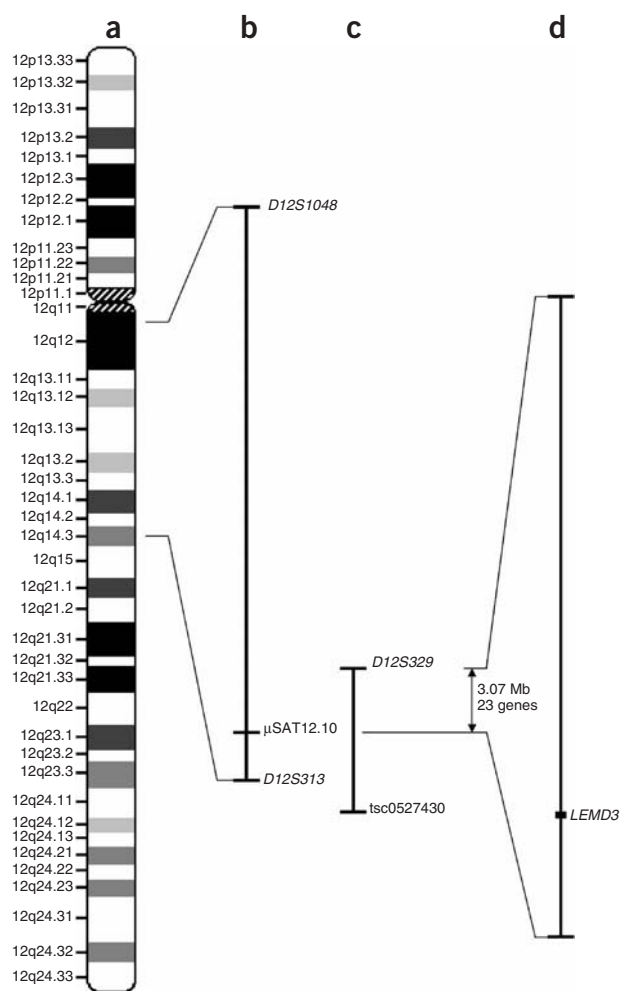


Figure 3 Ideogram of chromosome 12 showing the linkage interval, microdeletion and candidate region. (a) Ideogram of chromosome 12. (b) The 12q12–q14.3 linkage interval with indication of markers at the boundaries. (c) The microdeletion in individual G03-1858 in relation to the linkage interval. The region of interest is shown on the right (d), with *LEMD3* as the candidate gene.

with BOS, we found only the normal sequence, owing to nonsense-mediated decay of mRNA from the abnormal allele. Because bone specimens were not available, we could not investigate the possibility that a somatic mutation in osteoblasts could explain the spotty occurrence of bone lesions.

LEMD3 is an integral protein of the inner nuclear membrane¹⁶. It contains a nucleoplasmic N- and C-terminal domain and two transmembrane segments¹⁶. The N-terminal segment shares a conserved globular domain of ~40 amino acids with other inner nuclear membrane proteins, such as lamina-associated polypeptide 2 (LAP2) and emerlin¹⁶. The *X. laevis* ortholog of *LEMD3* (XMAN1) antagonizes BMP signaling. This antagonizing activity of XMAN1 resides in the C-terminal region that binds to Smad1, Smad5 and Smad8 (ref. 5).

To investigate whether *LEMD3* interacts with BMP receptor-activated and TGF β receptor-activated Smads, we carried out a yeast two-hybrid analysis using the C-terminal domain of *LEMD3* as prey. This analysis identified interactions between the C-terminal domain of *LEMD3* and the MH2 domains of Smad1 (BMP-specific)

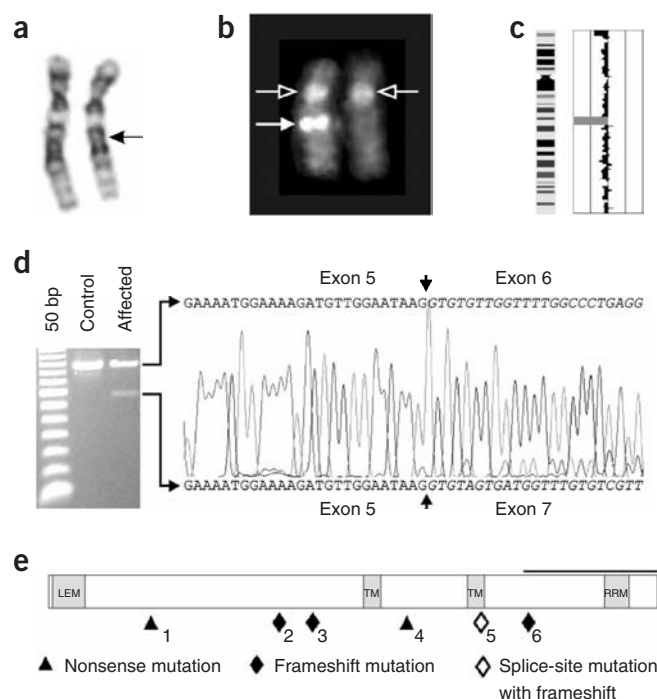


Figure 4 Overview of the cytogenetic and molecular defects found in affected individuals. (a) Partial karyotype from individual G03-1858 showing both chromosomes 12. The normal homolog is depicted on the left, and the homolog with the deletion (arrow) is shown on the right. (b) Metaphase FISH analysis with the BAC clone encompassing *LEMD3* (RP11-30506; filled arrow) and the centromeric 12 probe (open arrow), showing a microdeletion on the right homolog. (c) Results from the GeneChip Mapping 10K Array analysis for chromosome 12 of individual G03-1858. An ideogram of chromosome 12 is shown on the left; meta-analysis significance of the genetic copy-number variation of each SNP against the reference mean is shown on the right. The region with large negative values (bar to the left) indicates the presence of a microdeletion. (d) Effect of the *LEMD3* mutation in individual G03-2881. The electrophoretic analysis of an amplified cDNA fragment containing exons 4–8 shows the presence of a 146-bp shorter fragment as compared to the control. Partial nonsense-mediated decay is probably responsible for the weaker signal of this abnormal fragment. Sequence analysis shows skipping of exon 6 in the mutated allele (causing a frameshift with a premature stop codon in exon 7 at position 2,021; data not shown). (e) The positions of all *LEMD3* mutations identified in this study are shown below the structure of *LEMD3* (see also Table 1). Functional domains are indicated in gray: the LEM-containing N-terminal domain, the two transmembrane (TM) domains and the C-terminal domain with the RNA-recognition motif (RRM) motif. The black horizontal bar indicates the Smad-interacting part and BMP-antagonizing portion (as shown in XMAN1; ref. 5) of the C-terminal domain.

and Smad2 (TGF β -specific), suggesting that *LEMD3* is involved in both BMP and TGF β signaling (Fig. 5a).

We further investigated the role of *LEMD3* in both signaling pathways by overexpressing the protein in two different cell lines. In HEK293T cells, we measured the expression of several known target genes in basal conditions and after BMP4 stimulation by quantitative PCR (Q-PCR). Overexpression of *LEMD3* reduced the capacity of BMP4 to upregulate Smad6, Smad7, Id2 and Id3 (Fig. 5b). In HepG2 cells, we measured the response of TGF β using the activin-TGF β -responsive 3TP-Lux reporter in basal conditions and in the presence of a constitutively active receptor ALK4, which activates Smad2 and Smad3. Overexpression of *LEMD3* reduced the ALK4-induced

Table 1 *LEMD3* mutations

Number ^a	Affected family or individual	<i>LEMD3</i> mutation ^b
1	G03-1858	Total gene deletion
	G03-2882	457C→T
2	G03-1885	1033-1035delGGGinsC
3	Family C (G03-2457)	1185dupT
4	Family B (G02-1757)	1609C→T
5	G03-2881	1941+5delG
6	Family A (G02-1389)	2154dupA

^aThe numbers refer to the position of each mutation as shown in **Figure 4e**. ^bNumbering is according to cDNA sequence NM-014319.

activation of the 3TP-Lux reporter (**Fig. 5c**). These results indicate that *LEMD3* can antagonize both BMP and TGF β signaling in human cells.

Finally, we investigated the effect of the *LEMD3* mutations that we identified in affected individuals. We made three mutant *LEMD3* constructs containing the 1185dupT, 1609C→T and 2154dupA mutations, respectively. We carried out a luciferase assay in HEK293T cells transfected with a TGF β -responsive reporter and found that each mutant construct was unable to reduce TGF β signaling, unlike

the wild-type construct (**Fig. 5d**). The loss-of-function effect of these mutations most probably resulted from the absence of the Smad-interacting C-terminal domain in the truncated proteins, present on western blots. We therefore believe that the mutations found in this study are hypomorphic, either because of nonsense-mediated decay or because of the production of a truncated protein lacking the C-terminal domain. We measured by Q-PCR the expression of several target genes in skin fibroblasts from individual III-3 of family A with the 2154dupA mutation. This analysis confirmed that fibroblasts from this affected individual were haploinsufficient with respect to *LEMD3*. In addition, we found that expression of the gene *Id3* after TGF β stimulation was significantly higher in these fibroblasts than in controls (**Fig. 5e**). This is the first evidence to our knowledge that haploinsufficiency of *LEMD3* in human fibroblasts results in enhanced TGF β signaling with upregulation of target genes downstream in the pathway. We observed no significant differences between fibroblasts from the elastic-type nevus and normal skin of the affected individual in all conditions tested.

In conclusion, we found that loss-of-function mutations in *LEMD3* can result in osteopoikilosis, BOS and melorheostosis. The failure to detect a somatic mutation in *LEMD3* in affected skin fibroblasts from an individual with BOS and one with melorheostosis may suggest that

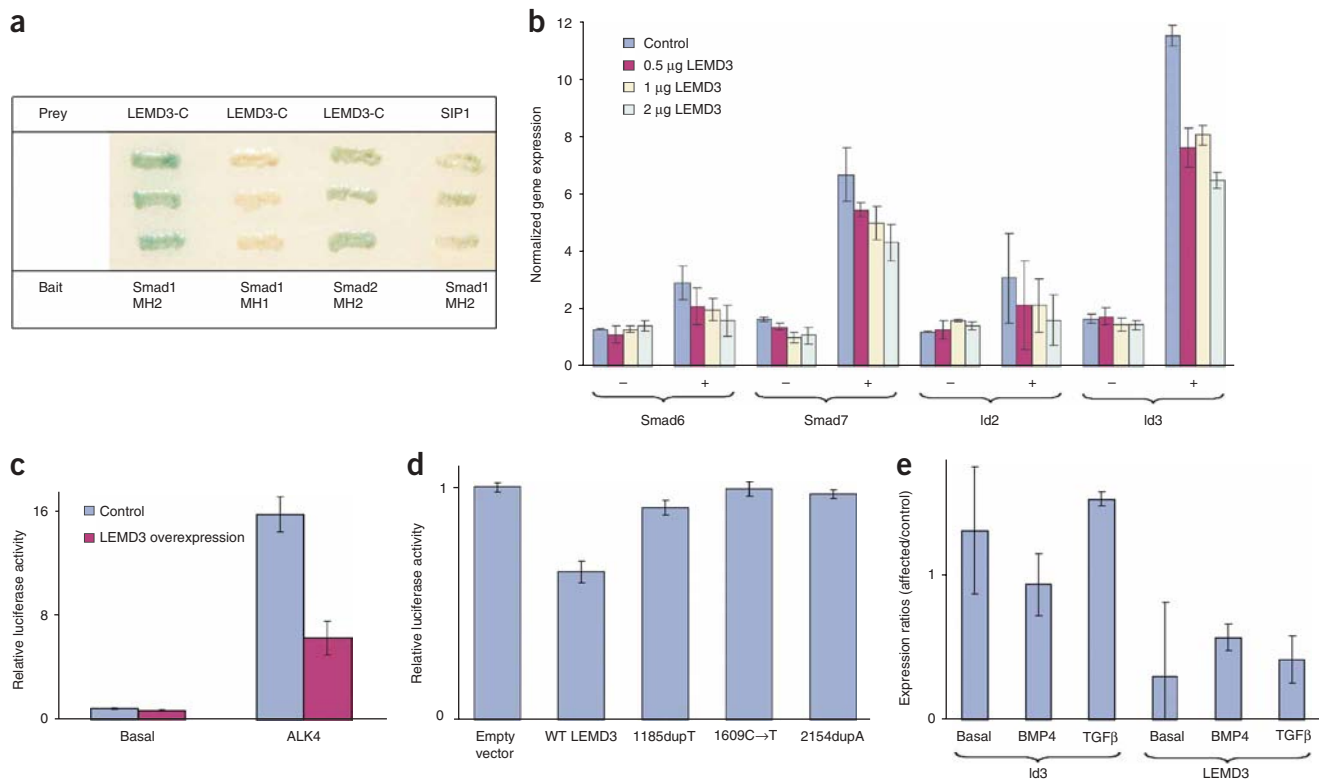


Figure 5 Analysis of normal and mutant *LEMD3* in BMP and TGF β signaling. **(a)** Yeast two-hybrid analysis shows that the C-terminal part of *LEMD3* (*LEMD3*-C) specifically interacts with the MH2 domain of Smad1 and Smad2. SIP1–Smad1 interaction is included as a positive control. Interaction was assessed by activation of the gene *MEL1* encoding α -galactosidase, visualized by blue staining of the yeast. **(b)** Normalized gene expression for four known target genes in basal (–) and BMP4-stimulated (+) HEK293T cells. Overexpression of *LEMD3* reduces the capacity of BMP4 to upregulate Smad6, Smad7, Id2 and Id3. **(c)** Relative luciferase activity from the 3TP-Lux reporter construct in HepG2 cells. Overexpression of *LEMD3* reduces activation of the 3TP-Lux reporter gene in the presence of the constitutively active ALK4 receptor. **(d)** Relative luciferase activity from the (CAGA)₁₂ reporter construct cotransfected with plasmids expressing different *LEMD3* mutants in HEK293T cells. Reduction of TGF β signaling is not observed for the different mutant *LEMD3* constructs. **(e)** Expression ratios between fibroblasts from an affected individual and control fibroblasts for *LEMD3* and *Id3* as measured by Q-PCR. The ratios for *Id3* in basal conditions and after BMP4 or TGF β stimulation are shown on the left. Significantly ($P < 0.05$) higher expression of *Id3* was observed after TGF β stimulation. The ratios for *LEMD3* are shown on the right and are indicative of haploinsufficiency in fibroblasts from the affected individual ($P < 0.05$).

other genetic factors contribute to the presence and distribution of skin and bone lesions in these disorders. In addition, we found evidence that *LEMD3* can antagonize both the BMP and TGF β signaling pathway. These results are in accordance with previous studies showing that genetic defects in both signaling pathways can result in hyperostotic bone disorders. Loss-of-function mutations in *SOST*, encoding the extracellular BMP antagonist sclerostin, lead to sclerosteosis^{17,18} (OMIM 269500), whereas activating missense mutations in *TGFBI* can result in Camurati-Engelmann disease^{19,20} (OMIM 131300). Finally, increased signaling in the TGF β pathway has been observed in other fibrotic skin disorders, such as scleroderma, and may therefore explain the skin lesions in individuals with BOS and melorheostosis^{21,22}.

METHODS

Material from affected individuals. We obtained appropriate informed consent from all subjects involved in the study. Family A is a three-generation family of Belgian origin. All affected individuals had osteopoikilosis. Two affected individuals (II-5 and III-3) also had skin manifestations of BOS. In individual III-3, skin biopsy samples were taken from a connective-tissue nevus on the left thigh and from normal skin on the right thigh. Light microscopy showed more numerous elastin fibers in the middle and deep dermis on the affected side. These fibers had a slightly granular and thickened appearance (Fig. 1b). Ultrastructurally, the elastin fibers were much thicker and more numerous in the affected skin than in the normal skin. They had a well-developed amorphous matrix with delicate peripheral microfibrils. Results of light microscopy and electron microscopy analysis were consistent with the diagnosis of connective-tissue nevus of the elastic-tissue type. Fibroblasts from both biopsies were cultured for molecular analysis.

Family B is another three-generation family of Belgian origin, previously reported², illustrating the co-occurrence of osteopoikilosis and melorheostosis in one family. Family C is a three-generation British family with autosomal dominant osteopoikilosis and skin lesions reminiscent of BOS. Individual G03-1885 is an Australian with BOS. Individuals G03-2881 and G03-2882 have osteopoikilosis and are both of Belgian origin but are unrelated. Individual G03-1858 had a history of prenatal and postnatal growth retardation and learning disability that was diagnosed as Russell-Silver syndrome in early childhood. Investigations for hypertension in infancy revealed ectopic kidneys and aberrant renal arteries. Evaluation at the age of 16 years showed proportionate short stature with height of 131.5 cm (50th centile for a 9-y-old girl), weight of 31.8 kg (50th centile for a 10-y-old girl) and head circumference of 49 cm (50th centile for a 3-y-old girl). Physical examination revealed a subtly dysmorphic face with synophrys, mild hypertelorism, broad and high nasal bridge, micrognathia and maxillary overbite. In addition, a diffuse hyperpigmentation spot was noted on the left thigh. Osteopoikilosis lesions were found on skeletal survey.

Linkage analysis. We used all autosomal markers from the Applied Biosystems Linkage mapping set version 2 for a genome-wide linkage analysis. We used an improved protocol from the Centre National de Génotypage for pooling an average of four markers per PCR to carry out all reactions. Additional markers were taken from the Marshfield map or designed based on the simple-tandem-repeat finder in the University of California Santa Cruz genome browser. We carried out genotyping on an Applied Biosystems Prism 3100 Genetic Analyzer running Genemapper v2.0 software. We used the MLINK program of the LINKAGE software package²³ to calculate two-point lod scores between the disease phenotype and each of the markers, assuming a dominant mode of inheritance with a penetrance of 0.95 and a disease allele frequency of 0.0001. The frequency of most marker alleles was set to 1/8. For markers with more than eight alleles in the pedigrees, the allele frequency was set to $1/n$, where n is the number of alleles.

Cytogenetic and FISH analysis. We carried out karyotyping in accordance with standard procedures. We analyzed metaphases at the 550-band level. We carried out FISH analysis as described²⁴ using BAC clone RP11-30506 from the 12q14.3 region.

GeneChip Mapping 10K Array analysis. We analyzed DNA from individual G03-1858 using the GeneChip Mapping 10K Array (Affymetrix) as described²⁵. We analyzed data with both the Affymetrix GCOS and GDAS software and the Affymetrix GeneChip Chromosome Copy Number Tool. We used copy number estimation (meta P value) to identify the microdeletion, the boundaries of which were defined by loss of heterozygosity. Results were visualized with arrayCGHbase (B. Menten *et al.*, unpublished data).

Sequence analysis. We amplified all exons by PCR using intronic primers and additional exonic primers for larger exons (primer sequences are available on request). We used a touchdown PCR program with an annealing temperature decreasing from 60 °C to 48 °C over 12 cycles, followed by 20 cycles with an annealing temperature of 48 °C. We sequenced PCR products using the BigDye v3.1 ET terminator cycle sequencing kit from Applied Biosystems. Sequencing reactions were loaded onto an Applied Biosystems Prism 3100 Genetic Analyzer and analyzed with Sequencing Analysis v3.7 and SeqScape v1.1 software (Applied Biosystems).

Yeast two-hybrid analysis. We cloned the C-terminal domain of human *LEMD3*, encoding amino acids 520–853, as a *PvuII-XhoI* fragment into the *SmaI* and *XhoI* restriction sites of the prey vector pAct2 (Clontech). We cotransformed yeast strain AH109 with this vector or a positive control (SIP1) prey vector with different Smad bait vectors²⁶, using the Yeastmaker yeast transformation system 2 (Clontech). We assayed activation of the gene *MEL1* by interacting hybrid proteins in yeast on Trp-Leu drop-out medium containing the chromogenic substrate X- α -Gal (Clontech).

Luciferase assay. We carried out transfections of HepG2 cells in triplicate in 24-well plates using Fugene (Roche). Each well was transfected with a total amount of 500 ng of plasmid DNA, including 50 ng of 3TP-Lux reporter, 50 ng of RSV-promoter-based *lacZ* reporter and combinations of 50 ng of the constitutively active ALK4 receptor with 100 ng of *LEMD3* or empty expression vector. We maintained a constant amount of DNA by adding pBluescript vector. Forty-eight hours after transfection, we assayed cell extracts for luciferase and β -galactosidase activities in accordance with the manufacturers' protocols (Promega and Clontech, respectively). We normalized the data by calculating the ratio of luciferase activity to β -galactosidase activity.

Q-PCR. We isolated RNA using RNeasy Mini Kit (Qiagen) and synthesized cDNA using SuperScript II Reverse Transcriptase Kit with random hexamer primers (Invitrogen) in a total volume of 20 μ l. We used 5 μ l of cDNA (1:10 dilution) in combination with the Q-PCR Core Kit for SYBR Green I (Eurogentec) and 250 nM gene-specific primers to carry out Q-PCR on a GeneAmp 5700 Sequence Detector (Applied Biosystems). The Q-PCR program consists of 40 cycles with 15 s at 95 °C and 1 min at 60 °C, followed by a dissociation run to determine melting curves. We carried out all reactions in duplicate and normalized them to the geometric mean of three stable reference genes (*GAPD*, *HPRT1* and *YWHAZ*)²⁷.

To investigate the effect of *LEMD3* on Smad signaling, we seeded HEK293T cells in 9-cm dishes and transfected them with a total of 2 μ g of *LEMD3* plasmid DNA (provided by H. Worman; Department of Medicine, College of Physicians and Surgeons, Columbia University, New York, New York, USA) using Fugene (Roche). Twenty-four hours after transfection, cells were serum-starved for 4 h and then stimulated with 5 ng ml⁻¹ of human recombinant BMP4 (R&D Systems) for 1 h followed by RNA extraction.

In the screening for differentially expressed genes, we used fibroblasts from a control individual and from normal and affected skin of affected individual III-3 (of family A). We grew the cells in 13.5-cm dishes to 80% confluency and then serum-starved them for 2 h. We collected cells after 1 h of treatment with 1 ng ml⁻¹ human recombinant TGF β 1 (R&D Systems) or with 5 ng ml⁻¹ of BMP4. Expression levels for all genes were determined in four independent experiments. Differential gene expression was considered significant when the difference was at least 50% and the 95% confidence interval of the mean expression ratio did not include 1 (equivalent with $P < 0.05$).

We analyzed the following genes (their ID numbers in the RTPPrimerDB database²⁸ are given in parentheses): *GAPD* (3), *HPRT1* (5), *YWHAZ* (9), *MMP2* (113), *CTGF* (596), *COL1A1* (1089), *COL3A1* (1090), *COL5A1* (1091), *FNI* (1092), *ELN* (1093), *ID1* (1094), *ID2* (1095), *ID3* (1096),

ID4 (1097), *LEMD3* (1098), *RUNX2* (1099), *SMAD6* (1100), *SMAD7* (1101) and *SERPINE1* (1102).

Analysis of mutant LEMD3 constructs. We modified the wild-type LEMD3 construct (provided by H. Worman; Department of Medicine, College of Physicians and Surgeons, Columbia University, New York, New York, USA) to contain the 1185dupT, 1609C→T or 2154dupA mutation using the QuickChange Site-Directed Mutagenesis kit (Stratagene) in accordance with the manufacturer's instructions. We plated HEK293T cells at a density of 4×10^5 cells per well in Dulbecco's modified Eagle medium plus (Invitrogen) and transferred them to Opti-MEM I (Invitrogen) after overnight adherence. We transiently transfected cells with 2 μ g of each plasmid and 0.5 μ g of the (CAGA)₁₂ TGF β -responsive reporter construct²⁹ in duplicate using Lipofectamine (Invitrogen). We also transfected cells with 20 ng of pRL-TK (Promega) to correct for transfection efficiency. Cells were serum-starved for 7 h before stimulation with 7 ng ml⁻¹ recombinant human TGF β (R&D systems). We lysed cells 24 h after transfection. We quantified activities of firefly and *Renilla* luciferase using the Dual-Luciferase Reporter Assay System (Promega).

URLs. Improved linkage analysis protocols from the Centre National de Génotypage are available at <http://www.cng.fr/>. The arrayCGHbase visualization tool and RTPrimerDB database are available at <http://medgen.ugent.be/arrayghbase/> and <http://medgen.ugent.be/rtpimerdb/>, respectively.

Note: Supplementary information is available on the Nature Genetics website.

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COMPETING INTERESTS STATEMENT

The authors declare that they have no competing financial interests.

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